

Synthesis of Novel Azapyridinomacrocyclic Trifunctional Chelatants for Biomolecules Labelling

Jennyfer YONG-SANG^{a,b,d}, Fabienne DIOURY^a, Clotilde FERROUD^a, Olivier MEILHAC^{b,c}, Emmanuelle JESTIN^d

^a Equipe Chimie Moléculaire du Laboratoire CMGPCE, Conservatoire national des arts et métiers (Cnam), 2 rue Conté, 75003, Paris, ys.jennyfer@live.fr

^b Groupe d'Etude sur l'Inflammation Chronique et l'Obésité (Geico) - EA 4526, Université de la Réunion - Campus du Moufia, Bât sciences, 15 avenue René Cassin, 97715, Saint Denis

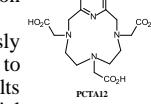
^c Inserm - U1148, Hôpital Bichat - secteur Claude Bernard, 46 rue Henri Huchard, 75018, Paris

^d Radiochimie et Imagerie du Petit Animal (Ripa), GIP Cyclotron Réunion Océan Indien (Cyroi), 2 rue Maxime Rivière, 97490, Sainte Clotilde

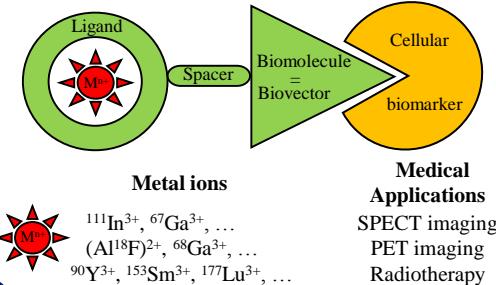
The coordination of metal ions is a method frequently used to label biomolecules as *biovectors* targeted to a specific cellular *biomarker*. Depending on the selected metal ion, the resulting targeted drug will find applications in medicine as diagnostic or therapeutic agents. For such purposes, multifunctional *ligands* are required in order to insure a strong and fast complexation of the metal ion on the one hand, and to enable a covalent grafting to the biomolecule on the other hand.

Polyamino-polycarboxylic compounds are known to give stable complexes avoiding any transchelation with endogenous cations as well as transmetalation with seric proteins, so that they are widely used in the medical area.¹

In order to prepare potential MRI contrast agents, we have previously synthesized such ligands based on the macrocyclic PCTA12-scaffold to immobilize Gd³⁺.²⁻⁵ The research presented here describes preliminary results of the synthesis of novel PCTA12-based trifunctional prochelators for potential multi-modal medical applications.

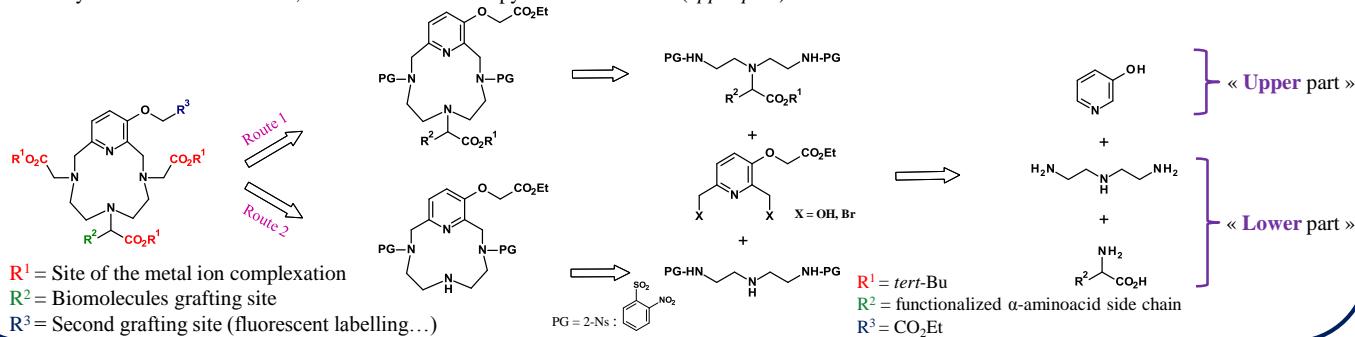


Labelled biomolecules as targeted drugs

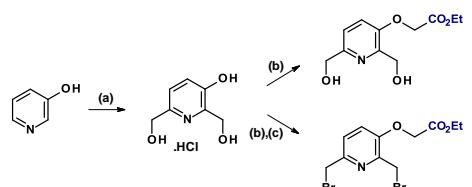


PCTA12-based trifunctional prochelators - Retrosynthesis

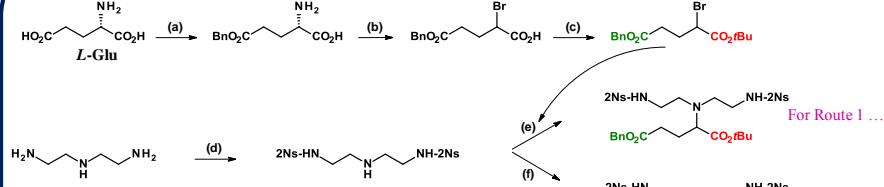
Routes 1 and 2 differ by the stage at which the part bearing the grafting site for the biomolecule and one of the three sites of the metal ion complexation is introduced. In route 1, it occurs later on a versatile macrocyclic synthon. In both routes, the macrocyclization step can be envisaged either *via* a S_N process or *via* a Fukuyama-Mitsunobu reaction, and involves the same pyridinic derivatives (*upper part*).



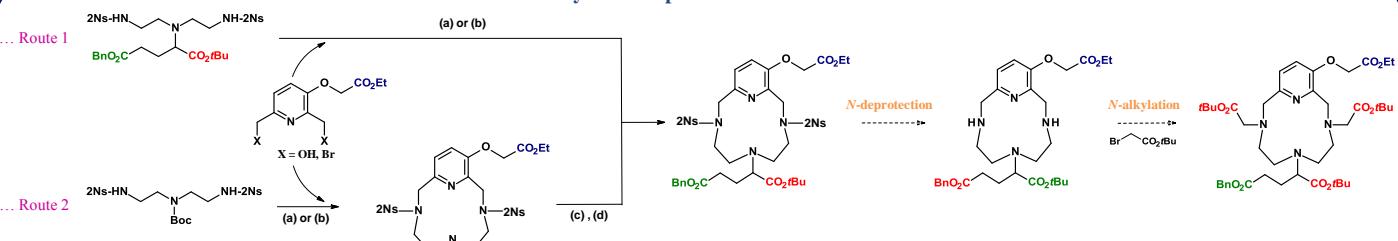
Synthesis of the « upper part »



Synthesis of the « lower part »



Macrocyclization procedures

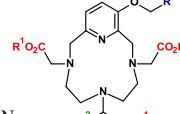


Reagents and conditions:
(a) For $X = \text{OH}$ (1 eq): PBu_3 (3 eq), DIAD (3 eq), THF_{amb} , rt, 24 h; 45 % (Route 1); 65 % (Route 2); (b) For $X = \text{Br}$ (1.2 eq): K_2CO_3 (4 eq), CH_3CN , reflux, 9h; 61 % (Route 1); 76 % (Route 2)
(c) TFA (27 eq), CH_2Cl_2 , rt, overnight; 97 % (d) $\text{C}_{16}\text{H}_{21}\text{BrO}_4$ (3 eq), K_2CO_3 (4 eq), KI (0.25 eq), $\text{CH}_3\text{CN}_{\text{amb}}$, reflux, 48h; 44 %

Perspectives

- Last two synthetic steps required (*N*-deprotection/*N*-alkylation) to obtain a first prochelator ($R^1 = \text{tert-Bu}$, $R^2 = (\text{CH}_2)_2\text{CO}_2\text{Bn}$, $R^3 = \text{CO}_2\text{Et}$)
- Orthogonal protective groups: cleavage tests
- Measurement of the stereochemistry of the prochelators prepared (chiral derivatization)
- Functional modulation of grafting sites R^2 , and R^3

$R^2 = (\text{CH}_2)_4\text{-NHCBz}; \dots$
 $R^3 = \text{COX-(CH}_2)_2\text{-NH-2Ns}$ with $X = \text{O}, \text{N}; \text{CH}_2\text{-NH-2Ns}; \dots$



1) L. Lattuada, A. Barge, G. Cravotto, G. Battista, L. Tei, *Chem. Soc. Rev.* 2011, 40, 3019-3049.

2) F. Diouri, C. Ferroud, A. Guy, M. Port, *Tetrahedron* 2009, 65, 7573-7579.

3) C. Ferroud, H. Borderies, E. Lasri, A. Guy, M. Port, *Tetrahedron* 2008, 64, 5972-5975.

4) F. Diouri, Y. Sambou, E. Guéné, M. Sabatou, C. Ferroud, A. Guy, M. Port, *Tetrahedron* 2007, 63, 204-214.

5) M. Port, I. Raynal, L. Vander Elst, R. N. Müller, F. Diouri, C. Ferroud, A. Guy, *Contrast Med. Mol. Imaging* 2006, 1, 121-127.